

**SOLIS  
BIODYNE**

## SOLIScript® RT cDNA synthesis KIT

Catalog number	Size (20 µl reactions)
06-35-0000S	50 rxn (sample)
06-35-00050	50 rxn
06-35-00200	200 rxn



**Store at  $-20^{\circ}\text{C}$   
upon receipt**

### Shipping:

At room temperature

### Batch Number and Expiry Date:

See vial

### Storage and Stability\*:

- Routine storage at  $-20^{\circ}\text{C}$  ( $-28^{\circ}\text{C}$  to  $-18^{\circ}\text{C}$ ) until Expiry Date
- Stable at  $4^{\circ}\text{C}$  ( $2^{\circ}\text{C}$  to  $8^{\circ}\text{C}$ ) for 3 months
- Stable at room temperature ( $15-25^{\circ}\text{C}$ ) for up to 1 month
- Freeze-thaw stability: 30 cycles

### Reaction setup:

At room temperature

Manufactured by Solis BioDyne, in compliance with the ISO 9001 and ISO 13485 certified Quality Management System.

## Kit content:

Components:
SOLIScript® Reverse Transcriptase
RiboGrip® RNase Inhibitor (220 U/μl)
10x RT Reaction Buffer with DTT
Oligo (dT) Primer (100 μM)
dNTP MIX (20 mM of each)
Water, nuclease free

## Product description:

- SOLIScript® is a chimeric genetically modified MMLV-based robust reverse transcriptase with reduced RNase H domain activity. It is an exceptionally stable enzyme due to a unique patented genetic modification in the polypeptide structure called the Stability TAG\*\*.
- The working temperature of enzyme ranges from 37°C – 60°C, with an optimum at 50°C. This property is beneficial for the detection of templates with complicated secondary structures.
- SOLIScript® is a fast enzyme with a reaction time of 5-30 minutes and is able to transcribe up to 8.9 kb RNA sequences.
- The enzyme is compatible with oligo (dT) and gene-specific primers, and is the best choice for direct 1-step qPCR solutions, due to excellent performance in the presence of common inhibitors.

## Applications:

- Gene expression analysis
- Disease diagnosis

- Genomic analysis
- Structural studies of mRNA

### Step-by-step guidelines:

1. Thaw the reagents at room temperature. Mix each reagent by gentle vortexing or pipetting up and down, then centrifuge briefly.

2. Mix the following components.

Component	Volume	Final conc.
Oligo (dT) primers (100 $\mu$ M)/ Gene specific primer <sup>1</sup>	1 $\mu$ l	5 $\mu$ M 0.1–1 $\mu$ M <sup>2</sup>
dNTP MIX (20 mM each)	0.5 $\mu$ l	500 $\mu$ M
10X RT Reaction Buffer with DTT	2 $\mu$ l	1x
SOLIScript <sup>®</sup> Reverse Transcriptase (200 U/ $\mu$ l)	1 $\mu$ l	10 U/ $\mu$ l
RiboGrip <sup>®</sup> RNase Inhibitor (220 U/ $\mu$ l) <sup>3</sup>	0.1 $\mu$ l	1.1 U/ $\mu$ l
Template RNA <sup>1</sup>	0.1–5 $\mu$ g	Variable
Nuclease-free water	up to 20 $\mu$ l	
Total	20 $\mu$ l	

<sup>1</sup> Not included in SOLIScript<sup>®</sup> cDNA synthesis KIT. Supplied by the end-user.

<sup>2</sup> Titrate primer concentration for optimal concentration.

<sup>3</sup> If there is a need for intermediate dilution, use any 1X RT, RT-(q)PCR, or RT-LAMP reaction buffer to do so.

### 3. Incubate the reaction mixture as follows:

Step	Temperature	Time
Reverse transcription	50°C	5 min <sup>1</sup>
Enzyme inactivation	85°C	5 min

<sup>1</sup> 5 min is sufficient for the synthesis of 1 kb amplicon. Optimize the reverse transcription step according to the troubleshooting guide.

#### Priming options:

- Oligo(dT) primers:
  - Specifically binds poly(A) tail of a eucaryotic mRNA (the total RNA pool has 1–2% of poly(A)+ RNA)
  - Generates full-length cDNA
- Gene specific primer:
  - Specific to the gene of interest
  - Increases specificity of the assay
  - Designing gene-specific primers complementary the to the 3' end of the target

#### Considerations while working with RNA:

- Avoid RNase contamination:
  - Wear protective clothing, gloves, etc.
  - Use sterile and RNase-free disposables
- Prior experiment store RNA samples at -70°C or below
- To protect RNA template use RiboGrip<sup>®</sup> RNase Inhibitor (provided with the kit)
- Thaw and keep RNA samples on ice during the experiment

## IMPORTANT:

cDNA synthesis reaction mixture components may inhibit PCR reaction. Don't use more than 10% of the cDNA reaction mixture in a final PCR/qPCR reaction.

### Troubleshooting guide

#### Low or no yield:

- Reaction conditions are not optimal – determine the optimal temperature by running a temperature gradient of 37–60°C (for example 37, 42, 50, 55, 60°C) for 15 min. When the optimal temperature is identified the optimal reaction time can be tested (5, 15, 30, and 60 min).
- Poor quality of template – check the template's purity and integrity. A260/280 and A260/230 ratios should be equal to or higher than 1.8 and 2.0 respectively.
- Primer design and/or concentration is not optimal – titrate primer concentration (final concentration range between 0.1–1  $\mu$ M). Use dedicated software, such as open-source Primer3 and NCBI Primer-BLAST to design target-specific primers.
- Reaction components are degraded – check the storage conditions and expiry date of the reagents; perform a positive control with template and/or reagents previously known to amplify.
- Inhibition of PCR/qPCR reaction - prepare a 5-fold dilution of the cDNA reaction mixture. Use up to 10% of the cDNA reaction mixture in a final (q)PCR reaction.

## Truncated cDNA:

- Poor quality of template - check the template's purity and integrity. Control for the presence of inhibitors, contaminants, degraded template RNA (old samples, degraded while purification, wrong storage conditions, *etc*).
- Sample has a high secondary structure or GC content – preincubate RNA at 65°C for 5 min, then chill on ice for 15 min.

## False positive result:

- Poor quality of template - genomic DNA contamination. To prevent amplification from genomic DNA, design primers spanning exon-exon junctions of the target mRNA, or primers that hybridize with sequences in consecutive exons flanking a long (e.g. 1 kb) intron. Revise sample preparation step. DNase treatment can prevent genomic DNA contamination.
- Components or disposables are contaminated – check the storage conditions of the reagents; perform a negative control with no template.
- Reaction mixture composition is not optimal - reduce the amount of DNA polymerase by 10%.

## Source:

Purified from an *E. coli* strain that carries an overproducing plasmid containing a SOLIScript<sup>®</sup> Reverse Transcriptase gene.

## Unit definition:

One unit is defined as the amount of enzyme that will incorporate 1 nmol of dTTP into an acid-precipitable material in 10 minutes at 37°C using poly(rA)•oligo(dT) as a template in a total reaction volume of 50 µl.

## Safety precautions:

Please refer to Safety Data Sheet for more information.

## Technical support:

Contact your sales representative for any questions or send an email to [support@solisbiodyne.com](mailto:support@solisbiodyne.com)

**For research use only. Not for diagnostic use.**

DS-06-35 v3. Revised: 13.12.2023

Reason for revision: Format of the Data Sheet is revised. Description and step-by-step guidelines are updated. A troubleshooting guide is added. Trademark and Permitted Use info is updated.

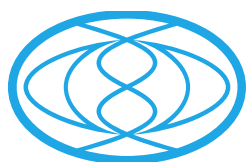
**\*Product stability** is assessed using routine QC assays and QC criteria set forth in the product specification and are intended to provide guidelines for shipping and storage conditions only. Customer or its designee shall be responsible for conducting all necessary stability testing applicable to their assay and/or QC criteria, and to comply with any applicable regulatory requirements or guidelines. Such stability testing shall include testing to validate the lead times for shipment, the shelf life of, and the product specifications applicable to shipment, storage and handling of the assay assembled and packed by the customer.

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**\*\*Covered by patent EP2501716, made following the methods of US Patent No 9,321,999.**

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